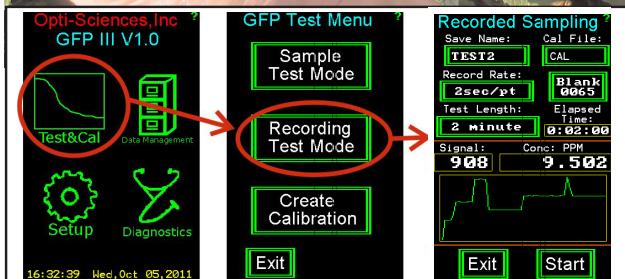




## Fluorescent Protein Meter

A hand-held device for the non-destructive determination of green fluorescent protein and other fluorescent protein compounds



- ◆ Non-destructive
- ◆ Hand-held, battery operated
- ◆ Accurate
- ◆ Touch screen, colour graphic display
- ◆ Large 2Gb internal memory
- ◆ Variety of other fluorescent compounds

### Non-destructive and battery portable

The GFP III Meter is a revolutionary new instrument, employing a modulated fluorescence technique for the non-destructive, quantitative measurements of Green Fluorescent Protein (GFP) in intact leaves.

Weighing only 275gms the GFP III Meter is a lightweight, hand-held, field portable instrument.

Programming and operation is easy, via a colour touch screen graphic display. Data and traces are also clearly presented on this display. The integral 2Gb data storage allows many thousands of experimental results to be saved, before downloading via a USB port.

Powered by two AA rechargeable batteries, the power efficient GFP III Meter will operate for up to 8 hours between charging.

A leaf clip is supplied as standard.

### GFP and other fluorescent proteins

Green Fluorescent Protein is the most commonly used genetic marker in gene expression studies, associated with genetically modified crop research. GFP is only detected, in a leaf sample, if the marker gene has been expressed by the plant.

Measurements are made using a 4mm fibre optic probe. The user may select to make single measurements or conduct automated continuous measurements per second or up to per minute.

The GFP III Meter provides researchers with a simple, accurate and fast means to study and differentiate high and low GFP expressing plants and transformed from non-transformed plants.

By simply changing the internal filter set, the GFP III Meter can be used for measuring other fluorescent compounds such as Chlorophyll, Rhodamine and Fluorescein.



### References for GFP Meter (all versions).

- Li1 J., Brunner A.M., Meilan R., Strauss S.H. (2009) Stability of transgenes in trees: expression of two reporter genes in poplar over three field seasons Tree Physiol (2009) 29: 299-312
- Li1 J., Brunner A.M., Meilan R., Strauss S.H. (2008) Matrix attachment region elements have small and variable effects on transgene expression and stability in field-grown Populus Article first published online: 7 SEP 2008 DOI: 10.1111/j.1467-7652.2008.00369.x Plant Biotechnology Journal Volume 6, Issue 9, pages 887896, December 2008
- Stuart M., Dignan C., McClary D., Golder Associates (NZ) Ltd, Dunedin, NZ, Ltd, Takapuna, NZ. (2008) Evaluation of marine response tools: Subtidal containment and treatment system MAF Biosecurity New Zealand Technical Paper No: 2010/20 Prepared for MAF Biosecurity New Zealand By ISBN 978-0-478-37081-2 (print) ISBN 978-0-478-37082-9 (online) ISSN 1179-5832 (print) ISSN 1179-5840 (online) December 2008
- Millwood R.J., Moon H. S., and Stewart C. N., (2008) Fluorescent Proteins in Transgenic Plants Reviews in Fluorescence 2008, Reviews in Fluorescence, Volume 2008, 387-403, DOI: 10.1007/978-1-4419-1260-2\_16
- Leckie B. M. and Stewart C.N., (2008) Agroinfiltration as a technique for rapid assays for evaluating candidate insect resistance transgenes in plants From the issue entitled "Special Issue: Plant biotechnology in support of the Millennium Goals (Volume I)" Plant Cell Reports Volume 30, Number 3, 325-334, DOI: 10.1007/s00299-010-0961-2
- Palumbo RE., and Veilleux R.E., (2007) Variation of GFP expression in diploid and tetraploid transgenic potato American Journal of Potato Research Volume 84, Number 5, 393-401, DOI: 10.1007/BF02987185
- Halfhill M.D., Millwood R.J. , Rufty T.W. , Weissinger A.K. and Stewart C.N. (2004) Spatial and temporal patterns of green fluorescent protein (GFP) fluorescence during leaf canopy development in transgenic oilseed rape, Brassica napus L. Plant Cell Reports Volume 22, Number 5, 338-343, DOI: 10.1007/s00299-003-0696-4
- Matthew D. Halfhill R., Millwood R.J., Stewart C.N., (2004) Green Fluorescent Protein Quantification in Whole Plants ,Reginald J. and C.Neal Transgenic Plants: Methods and Protocols Methods in Molecular Biology, 2004, Volume 286, IV, 215-225, DOI: 10.1385/1-59259-827-7:215
- Millwood R.J. , Halfhill M.D. , Harkins D. Russotti R., and Stewart, Jr.C.N. (2003) Instrumentation and Methodology for Quantifying GFP Fluorescence in Intact Plant Organs, UNC Greensboro, Greensboro, NC, 1NC State University, Raleigh, NC, and Opti-Sciences, Tyngsboro, MA, USA BioTechniques 34:638-643 (March 2003)

### Technical Specifications

**Measured parameters:** User programmable, can be calibrated to known concentrations or ratios. Multiple calibration tables may be generated.

**Sensitivity:** Depending on fluorescent compound for example Fluorescein:10ppb, Rhodamine 30ppb in pure water.

**Source:** LED selected for application in range 375nm-660nm. User changeable.

**Detector:** Solid state, high sensitivity detector. Band limiting filters set the detection range between 400nm-750nm. User changeable.

**Detection technique:** Modulated light digitally controlled to minimise background detection. Source and detector temperature compensation.

**Storage capacity:** 2 Gb internal non-volatile flash memory.

**Measurement modes:** Single point or continuous measurements from per second up to per minute.

**User interface:** 240 x 320 pixel colour touch screen graphic display with stylus.

**Measurement interface:** Bifurcated fiber optic probe (4mm) with 3mm diameter measurement circle.

**Output:** USB 1.1

**Operating temperature range:** 5°C-45°C

**Batteries:** 2 x rechargeable AA batteries. Two spare batteries also included.

**Auto off interval:** 4 minutes (with no key press or download)

**Dimensions:** 12 x 9 x 3cms

**Weight:** 275g



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